

Resistance of endoparasites to de-wormers

The definition of resistance is the ability of parasites to survive a treatment which is generally effective against a species and a stage of that species of parasite. Nowadays, resistance of parasites to de-wormers is considered to be a major issue in equine medicine. Therefore, de-wormers should be prescribed in a well-reasoned way so as to maintain their effectiveness, i.e to prevent the development of resistances. Resistance should not be considered on an individual basis, but on the basis of all the horses in a yard, as they share the same premises and pastures.

By **Marie DELERUE - Guillaume SALLE** - | 03.10.2016 |

Technical level   



What are the mechanisms leading to development of resistances ?

A parasite becomes resistant when its **genetic material is modified**. This **spontaneous mutation** is a natural phenomenon, and is not due to the parasite becoming accustomed to the de-wormer. It concerns a very small number of parasites among millions, but allows **the mutant parasites to persist after de-worming, to breed, and to transmit this resistant gene to the next generation**. This genetic mutation allows the parasites to resist a family of anthelmintic molecules which act in the same way. There are also **multiple resistances** which concern several families of molecule.

Systematic and frequent de-worming therefore leads to eliminating sensitive parasites, and revealing resistant parasites. Thus, it favours the **expansion of resistances**.

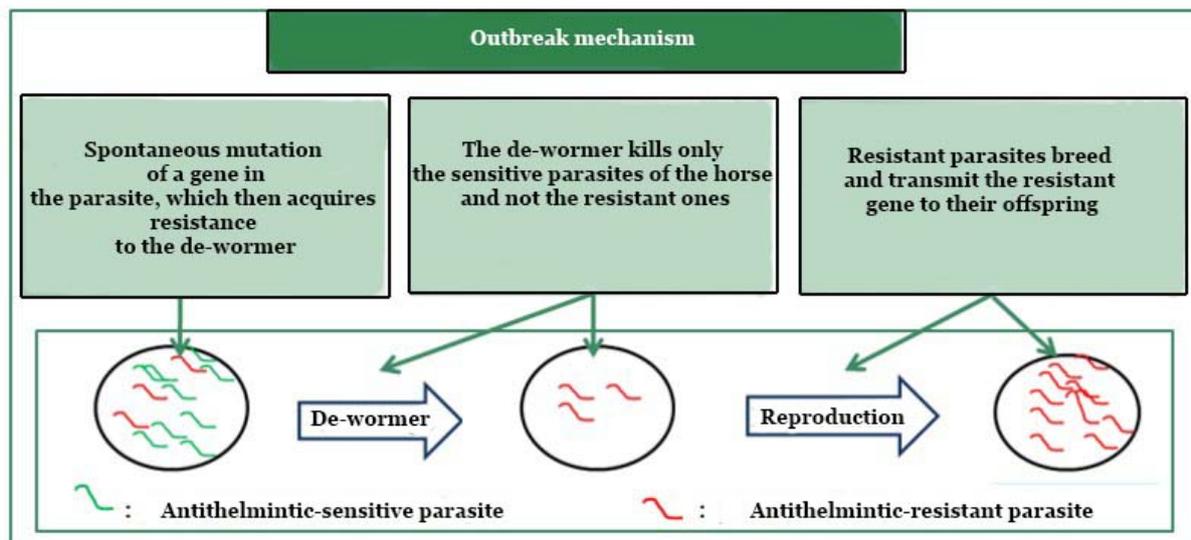


Diagram showing the mechanism of the outbreak of resistance to de-wormers

Assessment of resistances in France

In France, the most preoccupying resistances concern three families of de-wormer available on the market.

Resistance of small strongyles to benzidamoles

A recent study of 30 stud farms in 12 different departments confirmed the presence of resistant small strongyles to benzidamoles in 94 % of the studs tested. In studs where frequent de-worming was applied, resistance was the rule.

Resistance of small strongyles to Pyrantel

Of the 30 studs tested, resistance to pyrantel was suspected in 6 (20%) studs, and confirmed in 3 (10%).

Resistance of small strongyles to macrocyclic lactones

Macrocyclic lactones (ivermectin and moxidectin) demonstrate high effectiveness in most of the studs tested. However, in certain countries there has been a decrease of their efficiency.

Resistance of ascarids to macrocyclic lactones

In France, ivermectin-resistant ascarids have been found in Normandy

Assessment of antihelminthic-resistant small strongyles and ascarids in France in 2016		
	Parasites	
	Small strongyles (cyathostomins)	Ascarids
Resistance to benzimidazoles (fenbendazole, mebendazole)	Very high (in 94% of the farms tested)	Undocumented
Resistance to pyrantel	Moderate (in 10 % of the farms tested)	Undocumented
Resistance to macrocyclic lactones (ivermectin, moxidectin)	Rare (isolated cases)	Described in Normandy

The resistances described above mainly concern 2 parasites : small strongyles and ascarids. They are the most pathogenic in adult horses (small strongyles) and in young horses (ascarids and small strongyles). There are only 3 classes of antihelminthic molecule available on the market for equines, and there are no new molecules being developed at present. Consequently, it is of utmost importance to maintain the efficiency of these available molecules

What practices lead to an outbreak of resistance ?

- **Frequent systematic de-worming** of all horses in a yard : this induces high selection pressure, without leaving a refuge population, i.e parasites not subjected to de-worming;
- **Under dosing of the de-wormer** : administering an infra-therapeutical dose of antihelmintics will only eliminate the most sensitive parasites;
- **Using antihelmintic treatments which are not destined for equines** : their efficiency has not been tested on horses. Even in presence of a same molecule, the excipient, or the means of administration may be different, thereby generating the risk of under dosing;
- **Frequently introducing new horses** on the farm, they could potentially be infested with resistant parasites;
- **Fast rotation of different antihelmintic molecules** used, this can generate crossover resistances. Note : Not to be confused, antihelmintic molecules (there are only 3 available) and veterinary specialities which are numerous. One particular molecule may be present in several veterinary products;
- **Changing fields immediately after de-worming** : after treatment, horses shed only resistant parasites and eggs in their droppings. If the horses are then placed on a pasture with a low contamination level, the population of resistant parasites will colonise the pasture, and become dominant. If the horses are kept on the same pasture as prior to de-worming, the number of resistant parasites is diluted with sensitive parasites already on the pasture, and not selected by the de-wormer.

It is therefore of great importance **to respect the good practices of a well-reasoned de-worming program to avoid the outbreak of new resistances**

How can antihelmintic resistance of a parasite be measured ?

The presence of **antihelmintic resistant parasites** on a stud can be measured, specifically resistance of small strongyles to benzidamoles and pyrantel, where resistance is the most frequent. To determine whether a molecule is effective against small strongyles in a given yard, **a fecal egg shedding reduction test** is carried out. Overall resistance in a yard is tested, rather than resistance per individual horse. This is because, unless there is no contact between horses on the premises, they share the same parasites through the environment (fields, paddocks, stables.....). If there is a resistance it concerns all the horses in the yard.

To carry out this measurement, **two FECs need to be performed** (one **before de-worming**, and one **after treatment**).

First FEC carried out between day -1 and day -7

This first **FEC** gives a picture of **the initial number of eggs shed**, and enables to test only those horses with a high enough level of eggs shed so as to quantify the reduction after treatment.

It should be carried out on **a minimum of 12 horses** (so as to get a significant result) by **targeting potentially high shedders** (e.g young horses between 2 and 5 years old who have a lower immunity, and are therefore often more infested than adult horses).

De-worming - day 0

High and moderate shedding horses (i.e with an egg count of over 200 Epg) are de-wormed with the molecule to be tested.

Second FEC- Day 14

This **second FEC** is only performed for horses ranked high or moderate shedders in the first FEC, and for whom a treatment was implemented on day 0.

Thanks to a calculation comparing the egg count of the first FEC between day-1 and day-7, and the second FEC on Day 14, the efficacy of the molecule on the specific farm can be measured :

$$\% \text{ efficacy} = 100 \times [\text{epg (FEC 1)} - \text{epg (FEC 2)}] / \text{opg (FEC 1)}$$

Efficacy percentage

Below an efficacy percentage of 90 %, it is thought that there is **a resistance of small strongyles to the molecule tested**. Interpretation of the results remains delicate. The treating veterinarian will then decide, in accordance with the results on the given stud, whether to continue using the molecule or not.

Any **resistance is irreversible**, once detected and confirmed on a stud, the molecule will not become effective again, even when not used for a period of time.

Note : This measurement can be carried out for a single horse, but will not confirm the presence of resistant parasites in the herd. A resistance test can also be useful in foals : to measure the resistance of ascarids to ivermectin, and of small strongyles to pyrantel and benzimidoles.

When a new horse arrives in the yard, the test allows to determine the efficacy of the molecule used after treatment, and avoid resistant parasites coming into the stud through the new horse.

About our writers

- **Marie DELERUE** Docteur vétérinaire, ingénieur de développement IFCE
- **Guillaume SALLÉ** INRA - Unité Mixte de Recherche (UMR) 1282 - Infectiologie et Santé Publique - Nouzilly (37)

Bibliography

BOURDOISEAU G., 2015. La résistance aux anthelminthiques chez les équidés. *Le Nouveau Praticien vétérinaire équine*, vol. 11, pages 6-10.

GEURDEN T., BETSCH J.M., MAILLARD K.et coll., 2013. Determination of anthelmintic efficacy against equine cyathostomins and *Parascaris equorum* in France. *Equine Veterinary Education*, vol. 25, pages 304-307.

LAUGIER C., 2015. Etat des lieux de la résistance aux anthelminthiques chez les nématodes des équidés. *Le Nouveau Praticien vétérinaire équine*, vol. 11, pages 11-17.

LAUGIER C., SEVIN C., MENARD S.et coll., 2015. Prevalence of *Parascaris equorum* infection in foals on French stud farms and first report of ivermectin-resistant *P. equorum* populations in France. *Veterinary Parasitology*, vol. 188, pages 185-189.

NIELSEN M.K., MITTEL L., GRICE A., ERSKINE M., GRAVES E., VAALA W., TULLY R.C., FRENCH D.D., BOWMAN R. et KAPLAN R.M., 2013. AAEP, Parasite Control Guidelines.

SALLE G. et LAUGIER C., 2015. La détection de la résistance aux anthelminthiques chez les nématodes parasites des équidés. *Le Nouveau Praticien vétérinaire équine*, vol. 11, pages 18-22.

TRAVERSA D., CASTAGNA G., VON SAMSON-HIMMELSTJERNA G. et coll., 2012. Efficacy of major anthelmintics against horse cyathostomins in France. *Veterinary Parasitology*, vol. 188, pages 294-300.

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